Immunoinformatics: application of algorithmic approaches to solving immunological problems

Yana Safonova

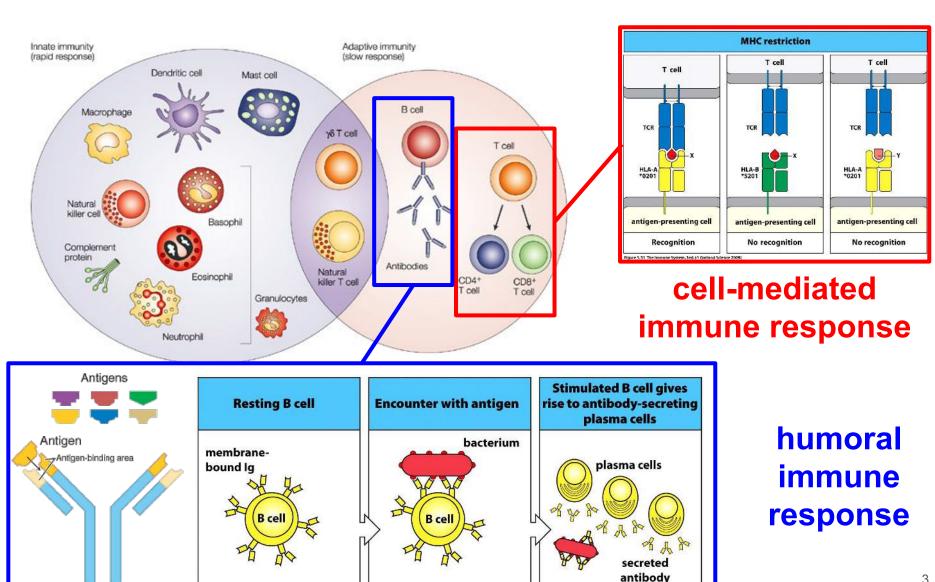
Center for Algorithmic Biotechnology St. Petersburg State University

Outline

- Introduction
- Repertoire construction problem
- Evolutionary analysis of antibodies
- Analysis of immune response dynamics
- Analysis of paired antibody repertoires & new biological insights from analysis of paired repertoires

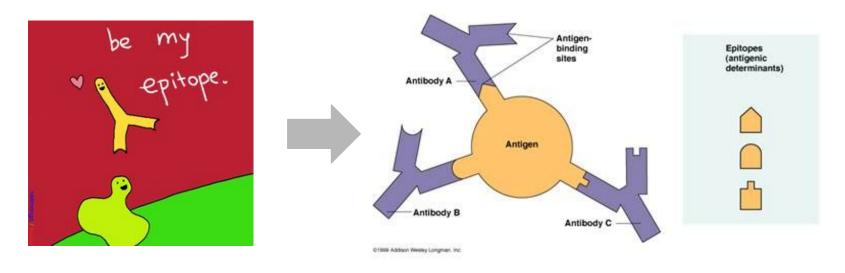
Innate & adaptive immune system

Figure 4.1 The Immune System, 3ed. (© Garland Science 2009)



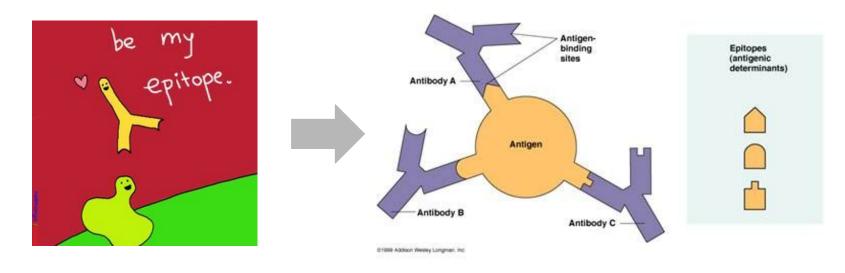


Antigen recognition

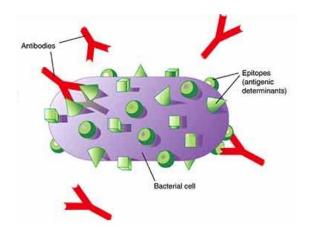


Antigen recognition

Antibody - antigen binding

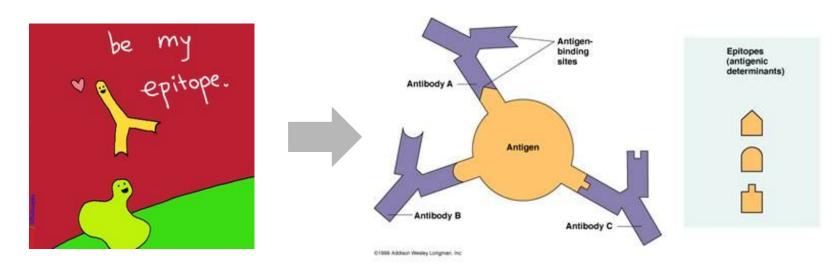


Antigen recognition

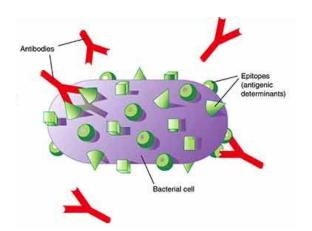


1. Antigen neutralization

Antibody - antigen binding

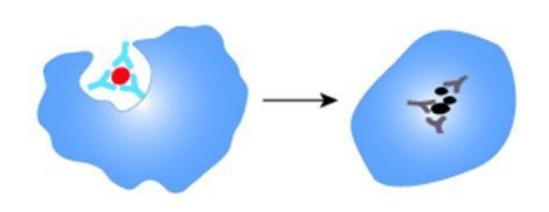


Antigen recognition



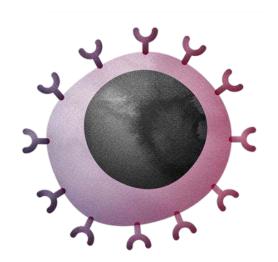
1. Antigen neutralization

Antibody - antigen binding

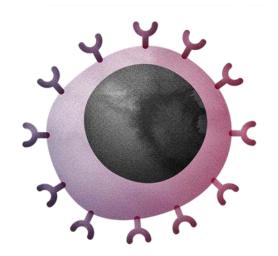


2. Destroying antigen by immune cells

Once you've met an antigen, your adaptive immune system never forgets it!



Once you've met an antigen, your adaptive immune system never forgets it!

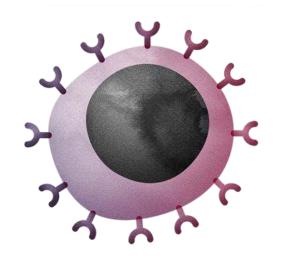


This principle is used for vaccine design:



Real antigens

Once you've met an antigen, your adaptive immune system never forgets it!



This principle is used for vaccine design:





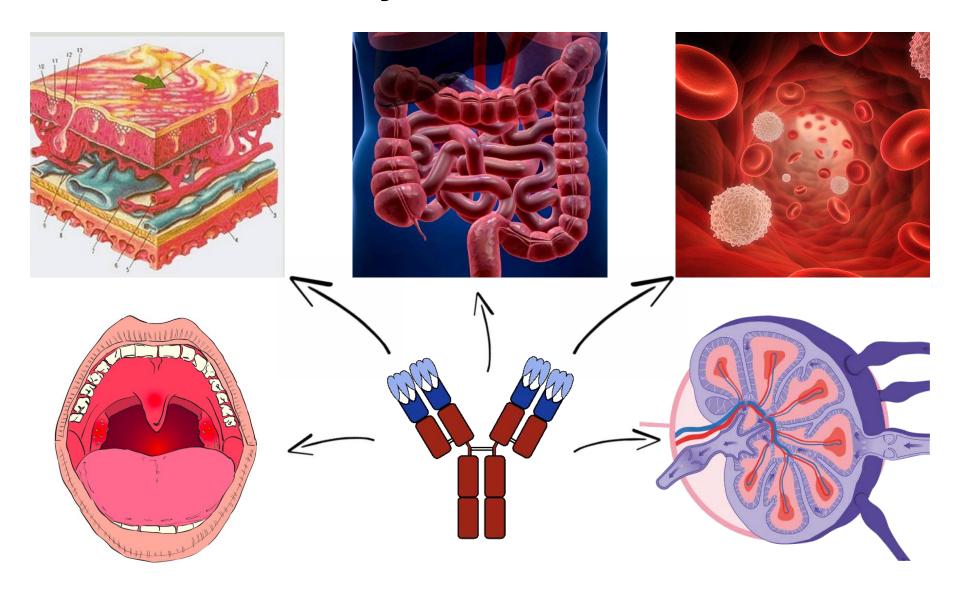


Vaccine



The Cow-Pock _ or _ the Wonderful Effects of the New Inoculation ! _ vide. the Publications of y Anti-Vaccine Society.

Where do antibody live?



Antibody repertoires

There is a **billion** of B-cells circulating in human blood at any given moment (out of 10¹⁸ estimated antibodies)

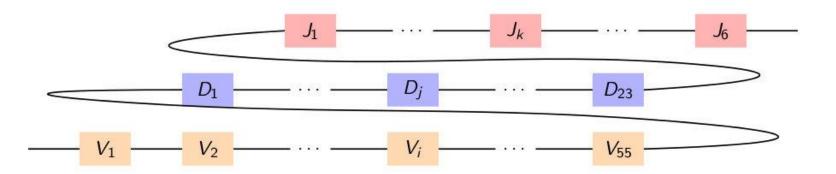


Analysis of concentrations of all antibodies in the organism (antibody repertoire) is a fundamental problem in immunology

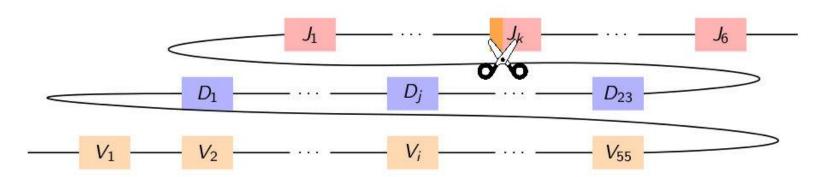
While generation of antibody repertoires provides a new avenue for antibody drug development, it remains unclear how to construct antibody repertoires from NGS data

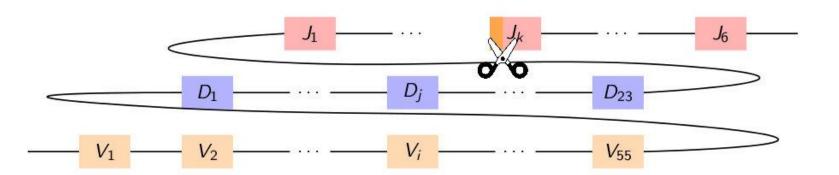
V(D)J recombination

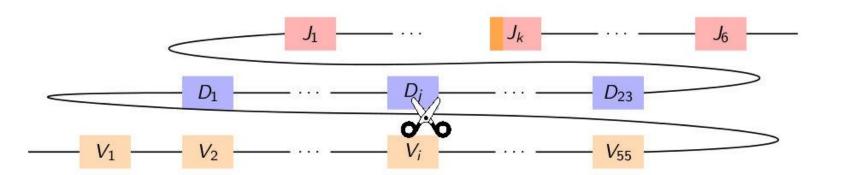
Antibodies are produced by **B-cells**, each with unique genome:

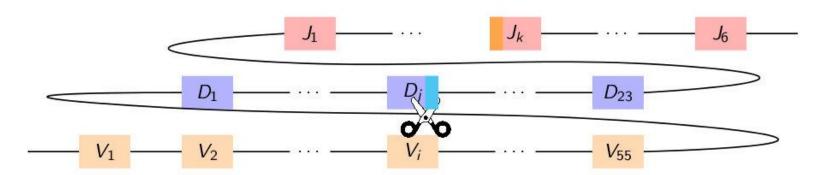


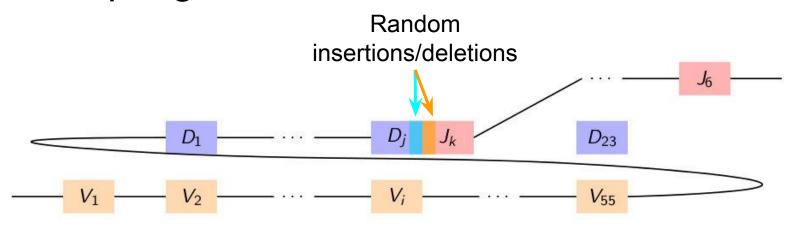
IGH locus in human genome (1 MB length)

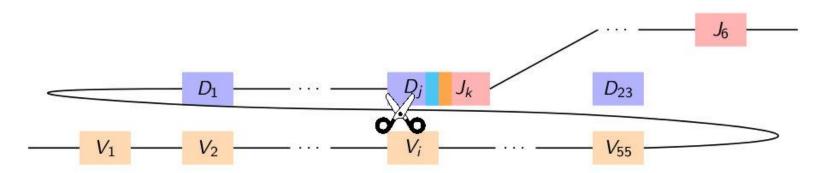


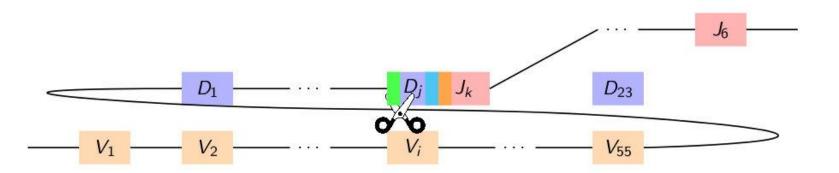


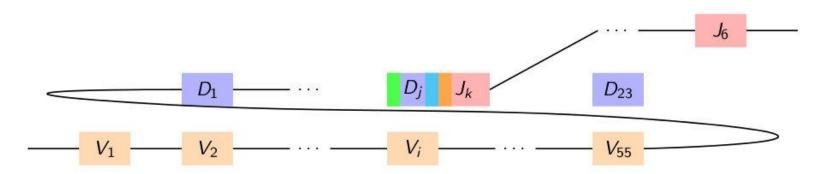


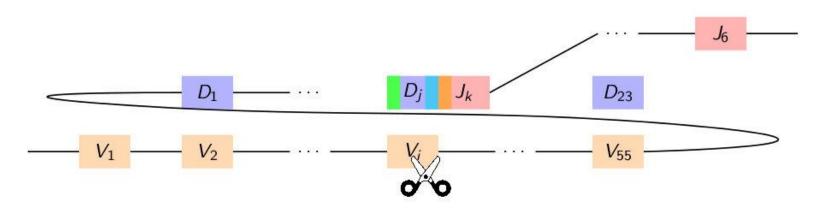


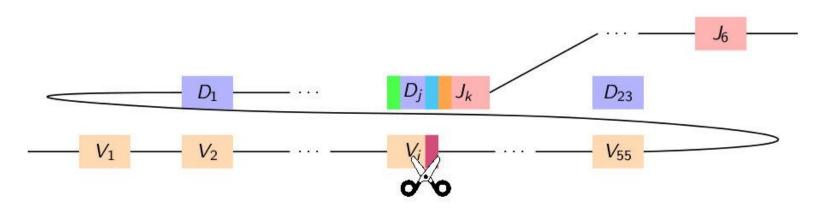


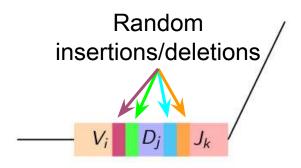




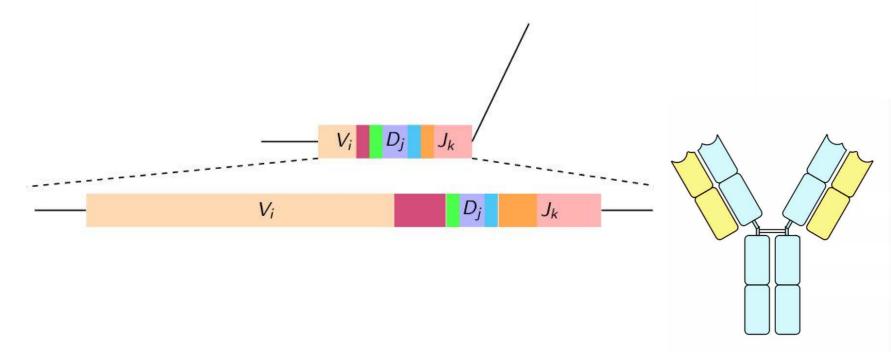






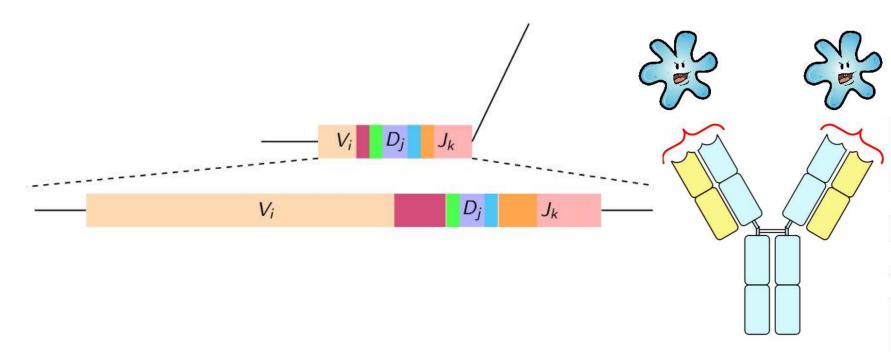


Somatic recombination results in unique immunoglobulins genes encoding amino acid sequence of *antibodies*



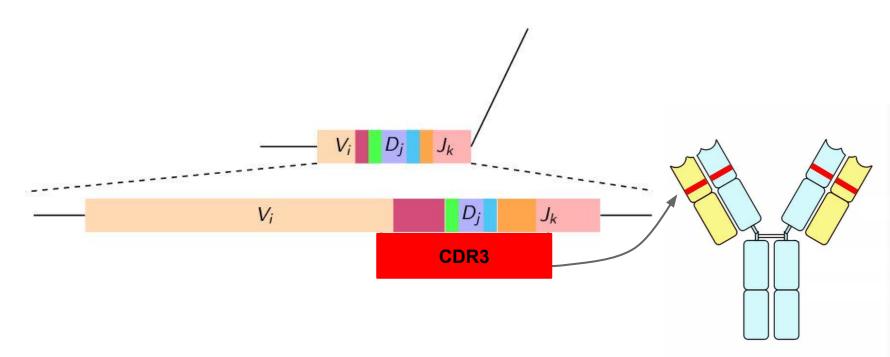
Antibody versus antigen

An antibody recognizes a foreign agent (antigen) using its antigen-binding site



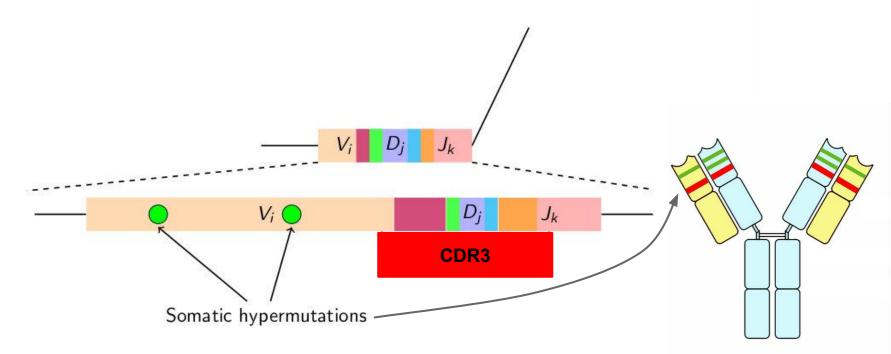
Antigen binding site in antibody

The most diverged part of antigen-binding site is complementarity determining region 3 (CDR3)

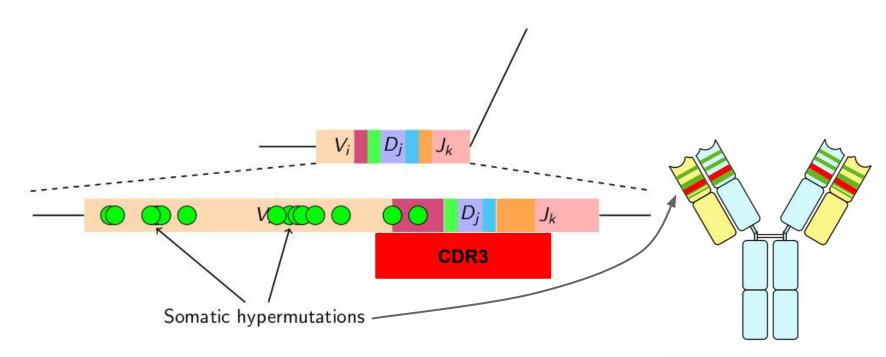


Somatic hypermutations

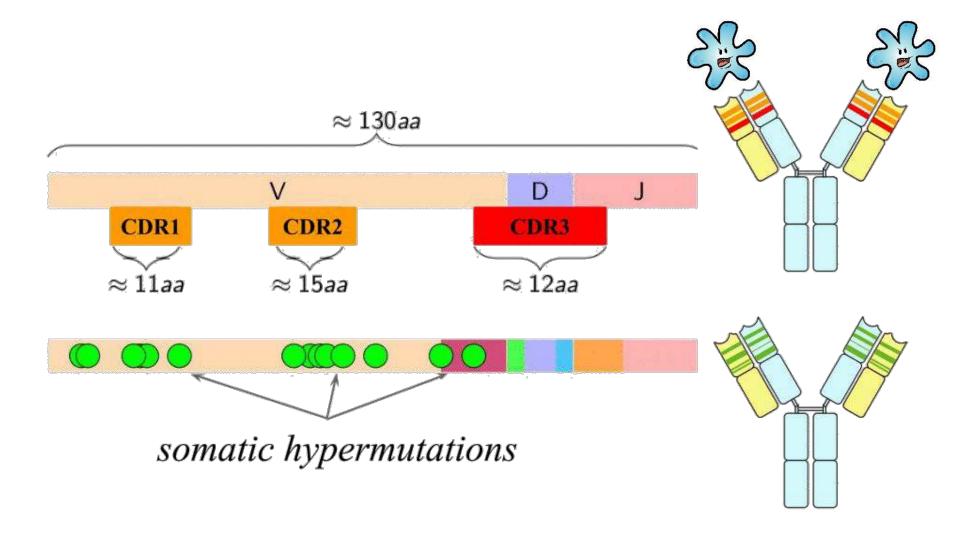
Further optimization of antibody affinity is achieved through somatic hypermutations



...many somatic hypermutations



Architecture of antibodies



VDJ classification problem. Given an antibody generated from a *known set* of V, D, and J segments, identify what specific V, D, and J segments generated this antibody



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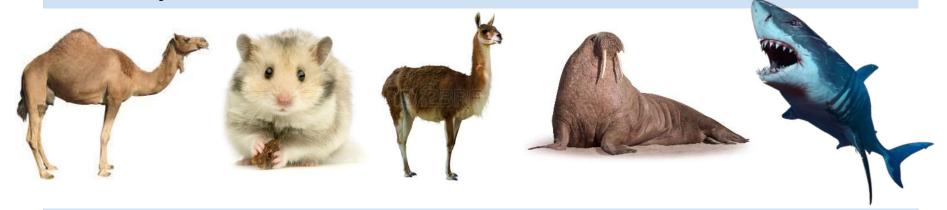


VDJ classification problem. Given an antibody generated from a *known set* of V, D, and J segments, identify what specific V, D, and J segments generated this antibody



Important model organisms in immunology with still unknown sets of V, D, and J segments

VDJ classification problem. Given an antibody generated from a *known set* of V, D, and J segments, identify what specific V, D, and J segments generated this antibody



VDJ reconstruction problem. Given a set (millions) of antibodies generated from an *unknown* set of V, D, and J segments, reconstruct these sets

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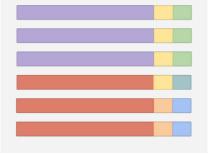
Roche 454 (2005)

low coverage

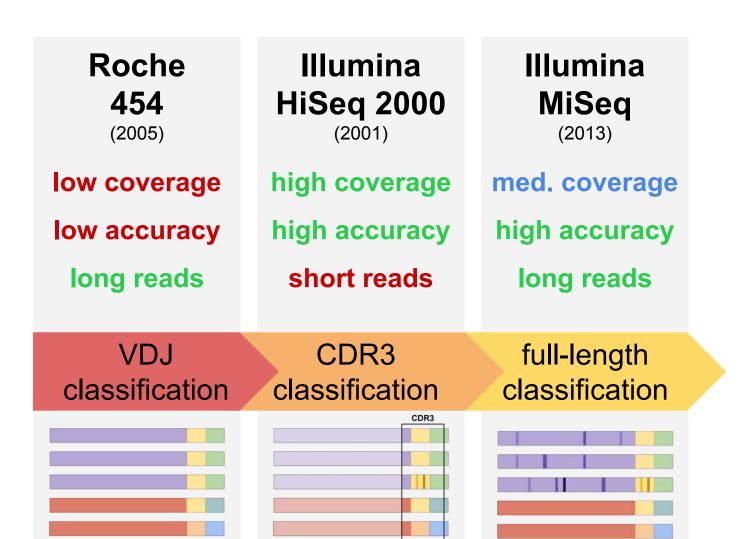
low accuracy

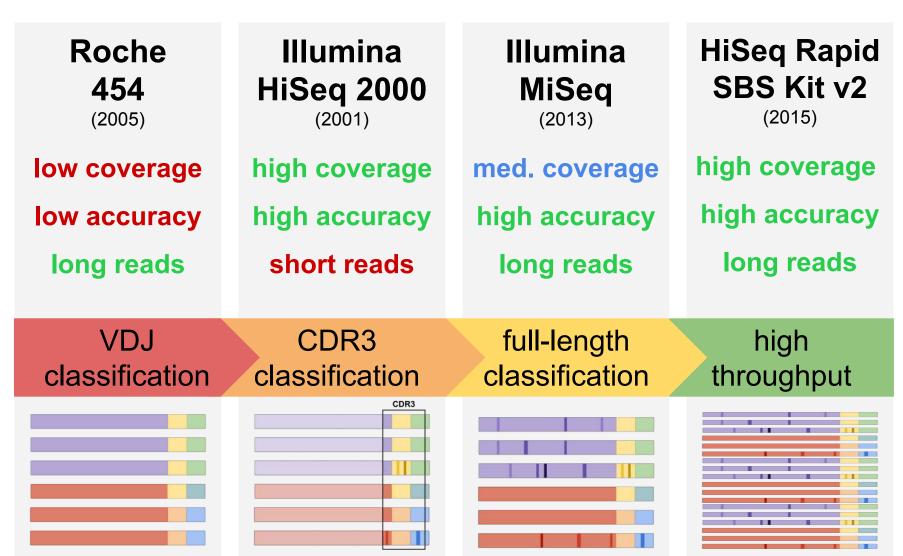
long reads

VDJ classification



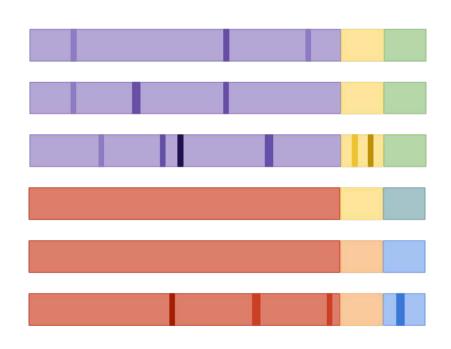
Roche Illumina 454 **HiSeq 2000** (2001)(2005)low coverage high coverage low accuracy high accuracy long reads short reads **VDJ** CDR3 classification classification





Full-length antibody classification (repertoire construction)

In contrast to well-studied VDJ and CDR3 classification, full-length antibody classification takes into account the entire variable region of antibody

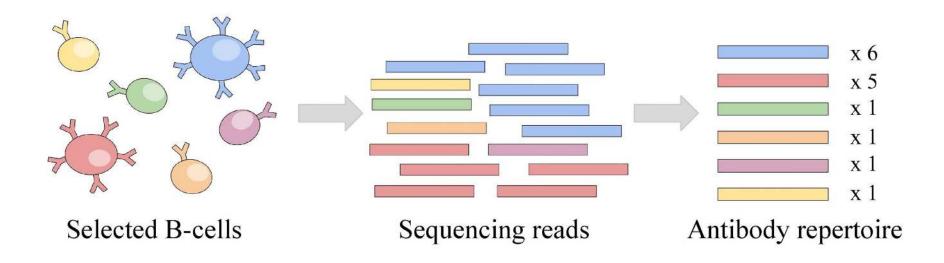


MiGEC: Shugay et al., *Nat Methods*, 2014 **MiXCR**: Bolotin et al., *Nat Methods*, 2015

IMSEQ: Kuchenbecker et al., Bioinformatics, 2015

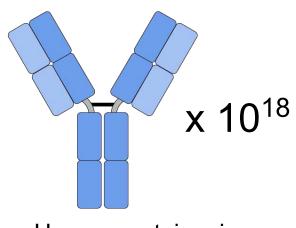
IgRepertoireConstructor: Safonova et al., Bioinformatics, 2015

Repertoire construction problem



- Giant read clustering problem
- Giant error correction problem

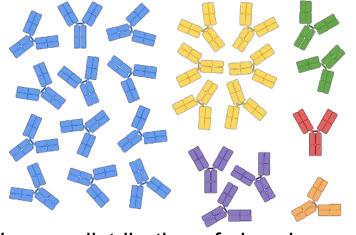
What makes this clustering problem difficult?



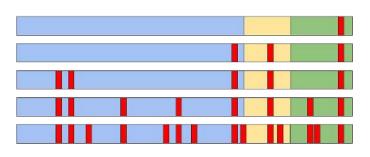
Huge repertoire size



High repetitiveness



Uneven distribution of abundances



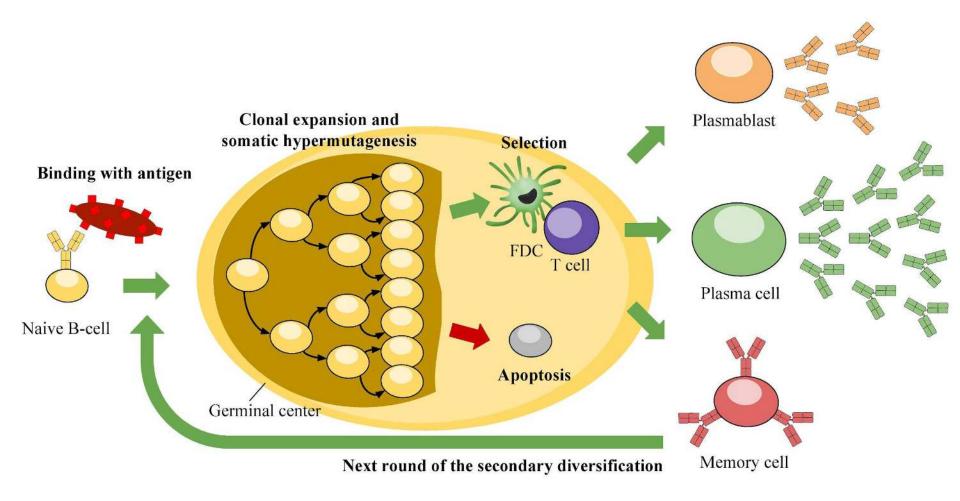
High mutation rate

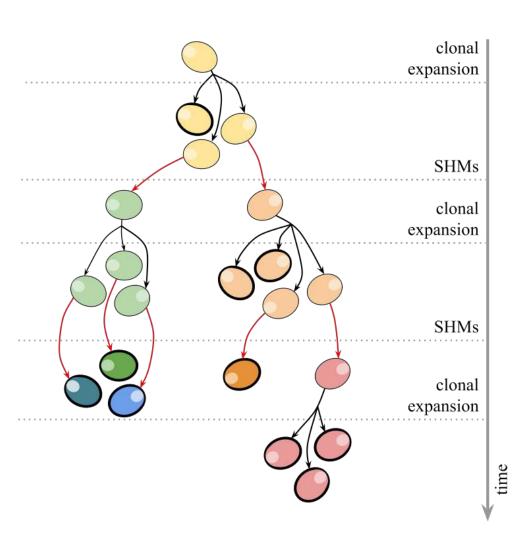
- Global coverage threshold cannot be used for error correction
- Sequencing errors often look like natural variations

Outline

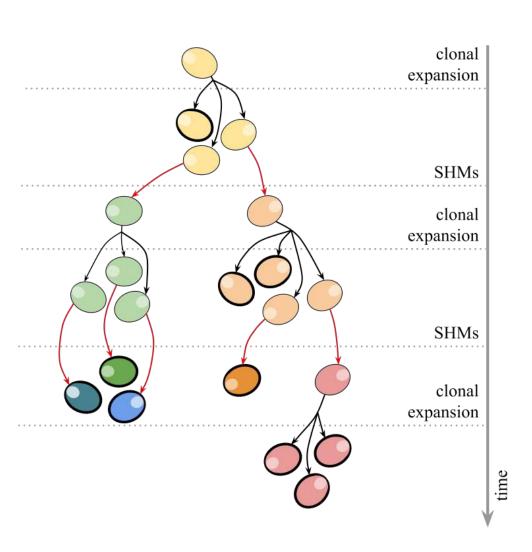
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Secondary diversification of antibodies

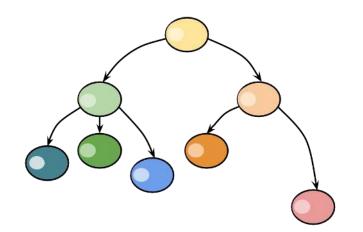


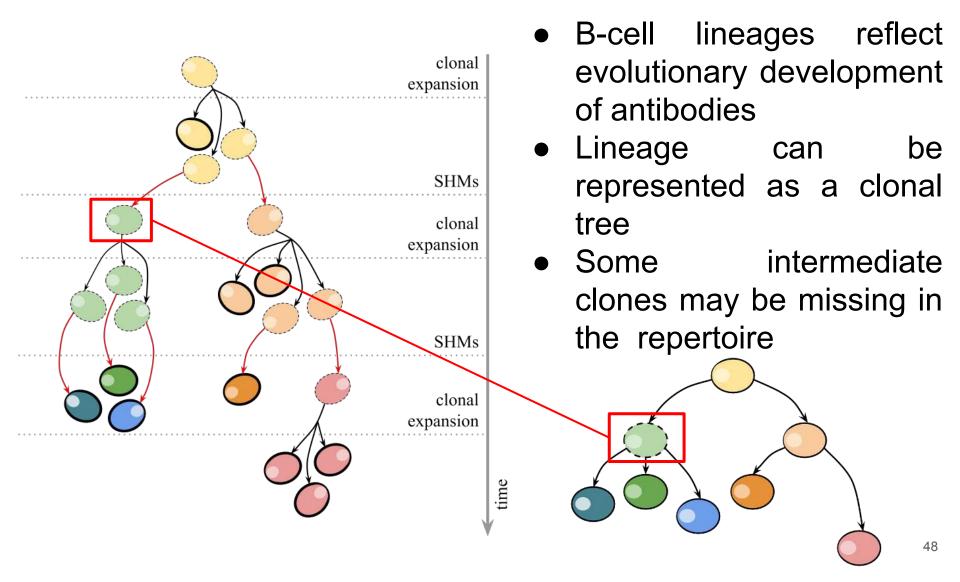


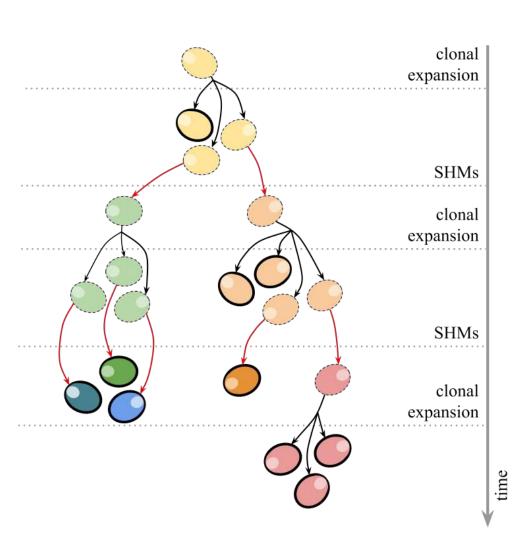
 B-cell lineages reflect evolutionary development of antibodies

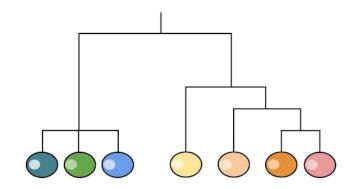


- B-cell lineages reflect evolutionary development of antibodies
- Lineage can be represented as a clonal tree

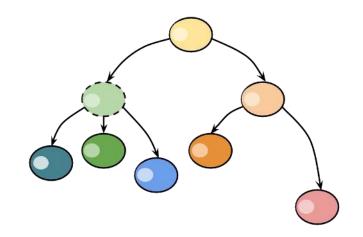




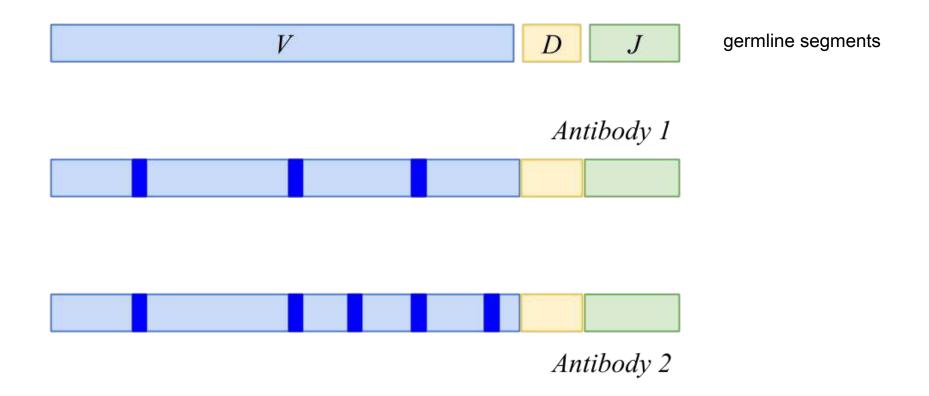




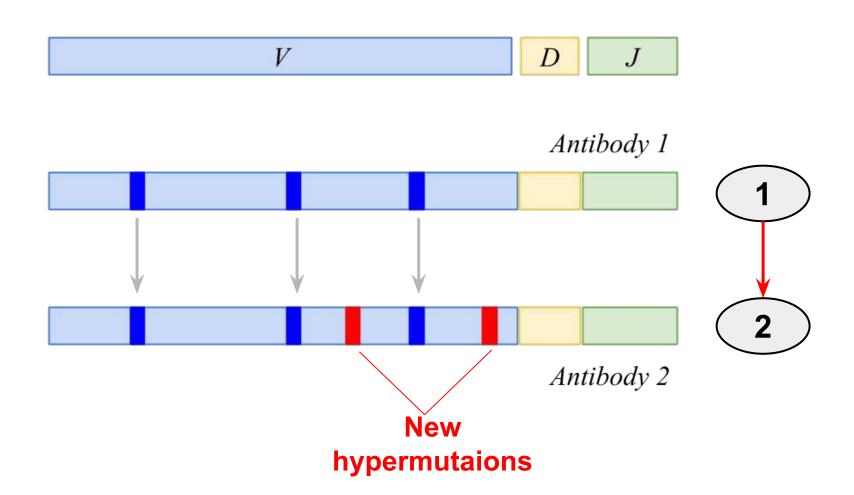
Standard phylogenetic algorithms assume that all species are represented by leaves and should be adapted for clonal trees



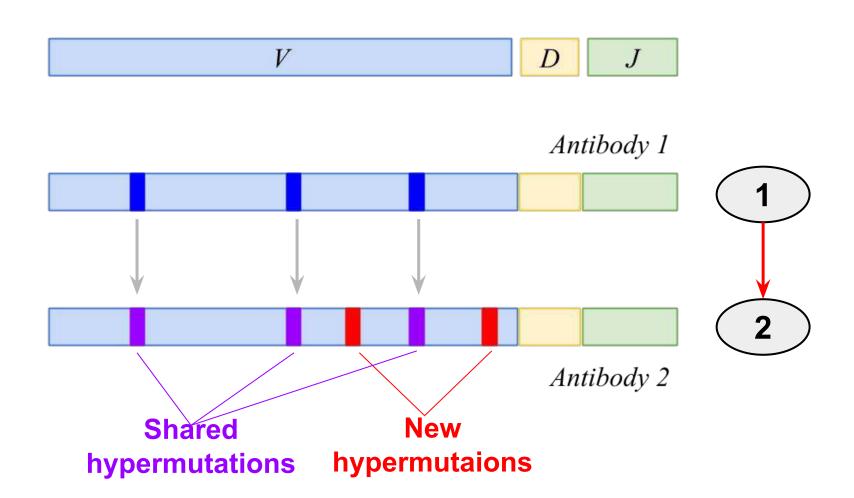
Who is the ancestor here?



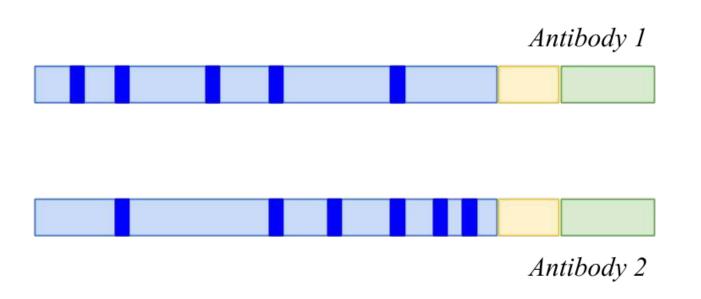
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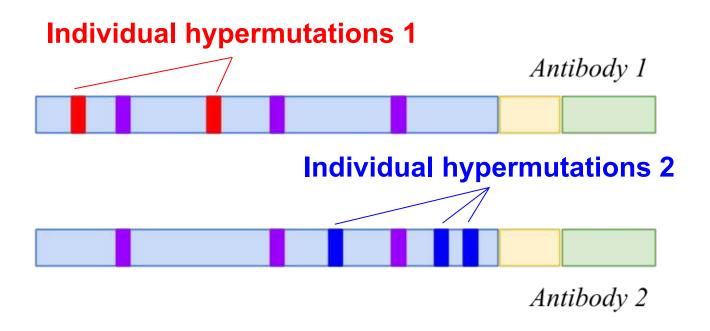
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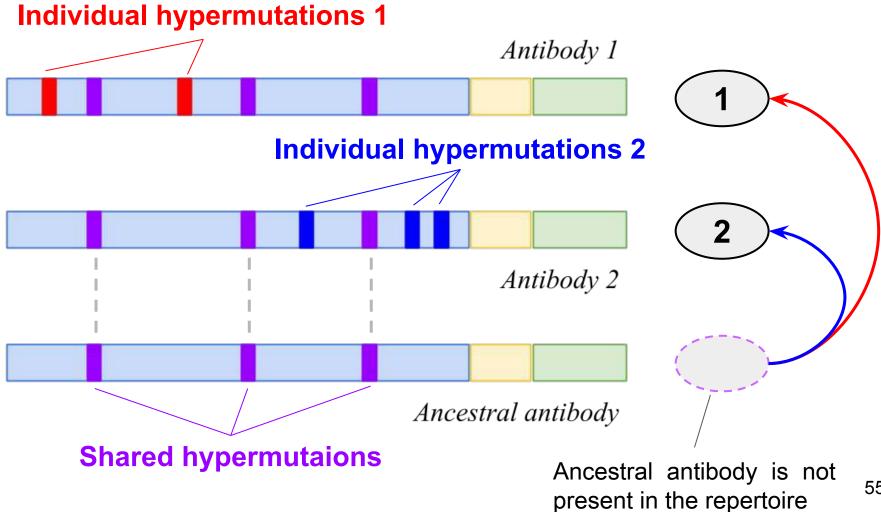
Another example: who is the ancestor here?



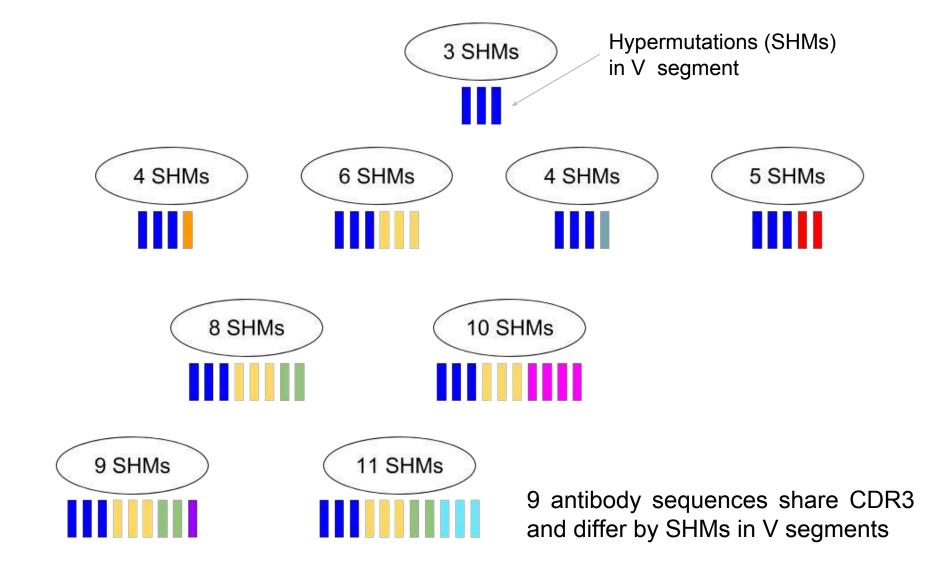
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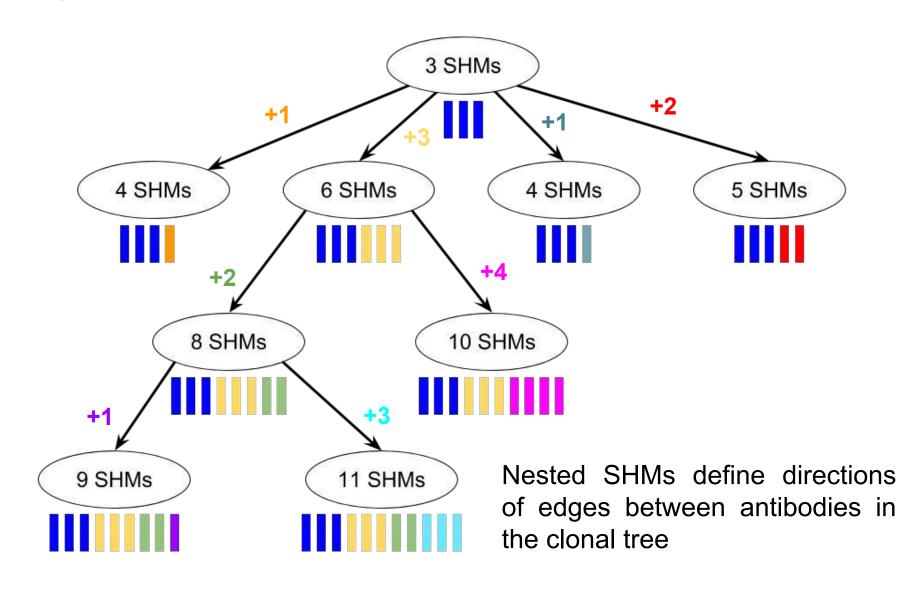
Ancestral antibody may be missing...



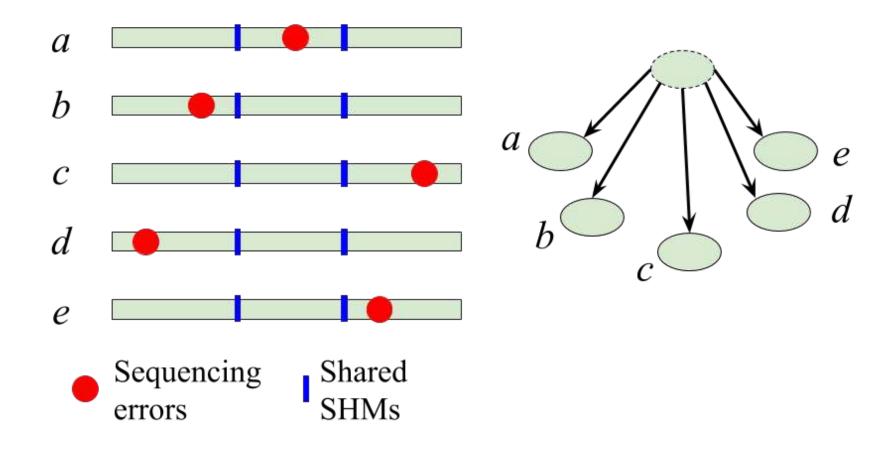
What is the evolutionary tree?



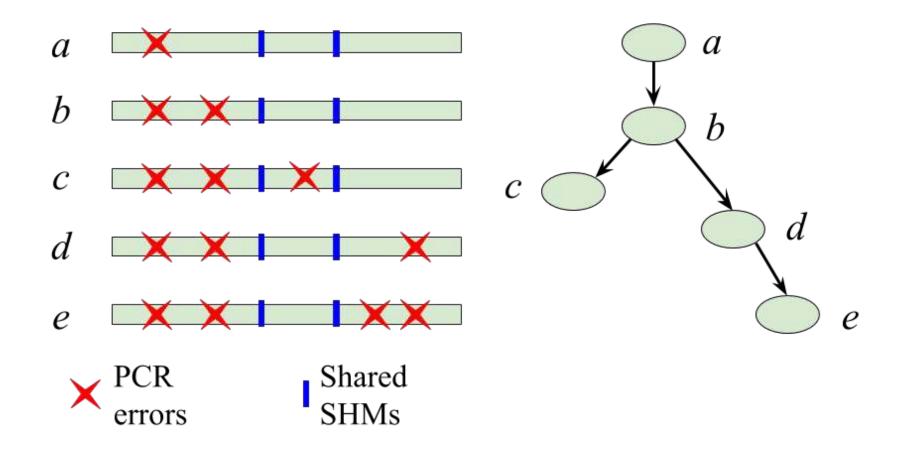
Any tree reconstruction approach will work



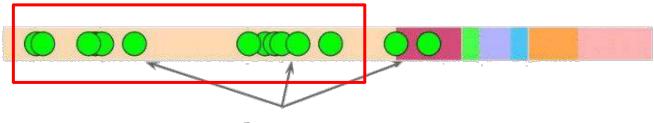
Repertoire construction step is very important for clonal analysis!



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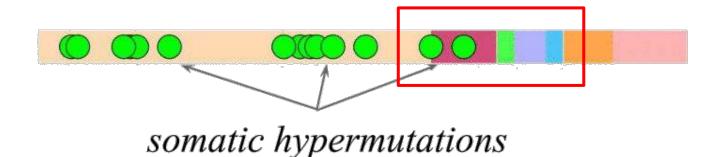
SHMs in V segments are easy to find



somatic hypermutations

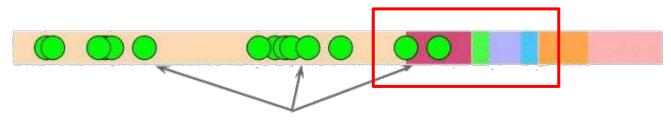
 One can easily identify mutations in the V segment using alignment against the **template** (germline V segment)

SHMs in CDR3 are difficult to identify



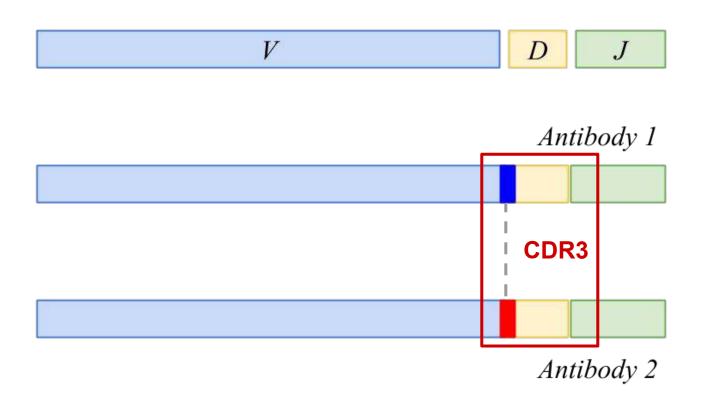
- One can easily identify mutations in the V segment using alignment against the **template** (germline V segment)
- But there is no template for CDR3!

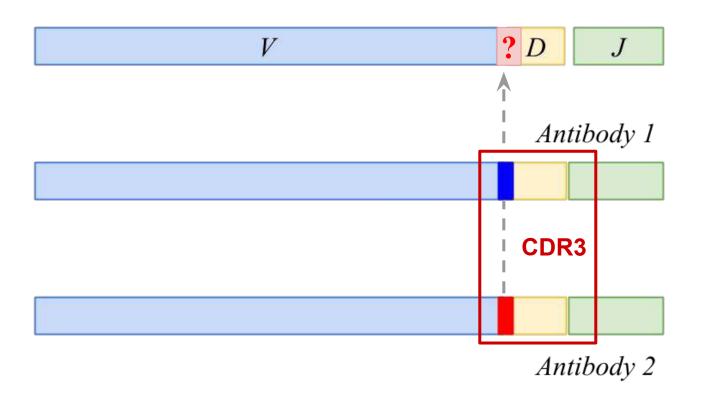
SHMs in CDR3 are difficult to identify

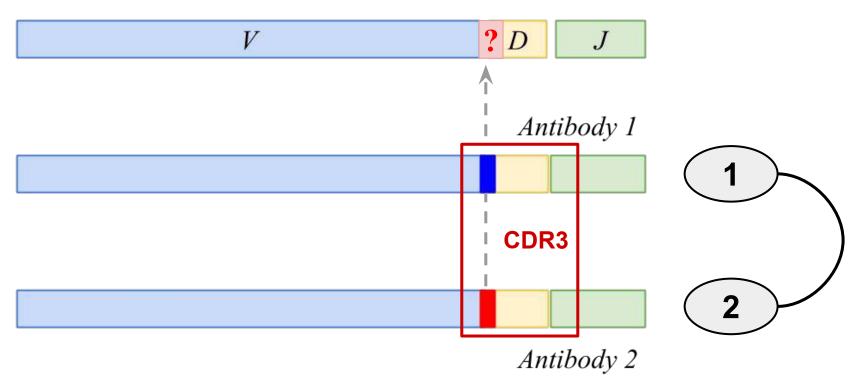


somatic hypermutations

- One can easily identify mutations in the V segment using alignment against the **template** (germline V segment)
- But there is no template for CDR3!
 - deletions in gene segments
 - non-genomic VD and DJ insertions
 - addition of palindromes

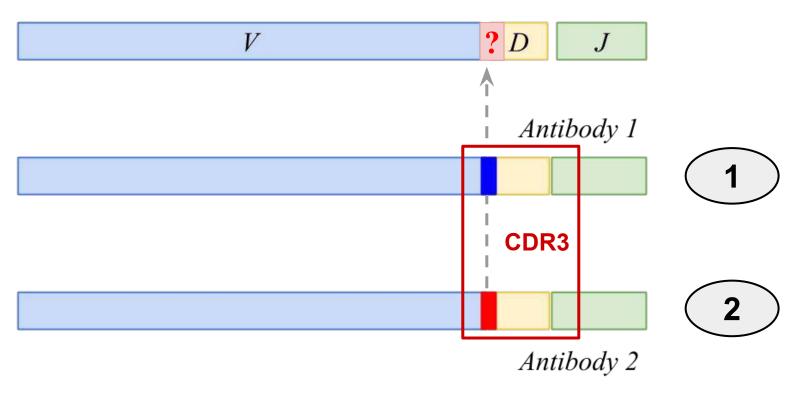






Information about VDJ scenarios allows us to make the a choice:

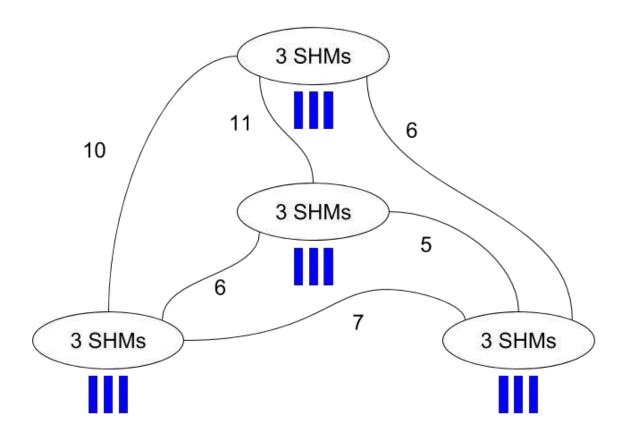
Antibodies 1 and 2 belong to the same lineage



Information about VDJ scenarios allows us to make the right choice:

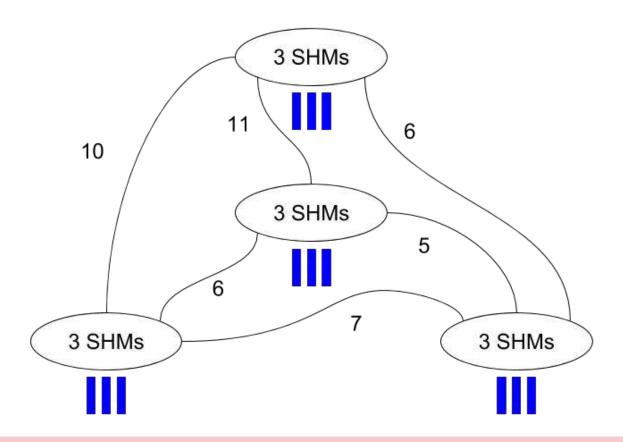
- Antibodies 1 and 2 belong to the same lineage
- Antibodies 1 and 2 are not related

Another puzzle



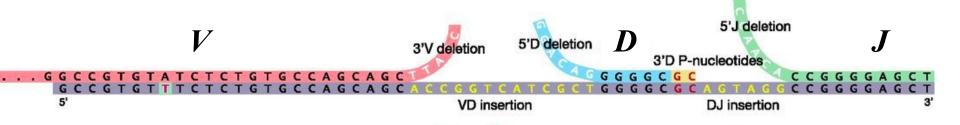
4 antibodies share SHMs in V segments but differ in CDR3s

Another puzzle



- It is unclear how to select direction between two similar CDR3s
- It is unclear whether two similar CDR3s belong to a single clonal tree or not

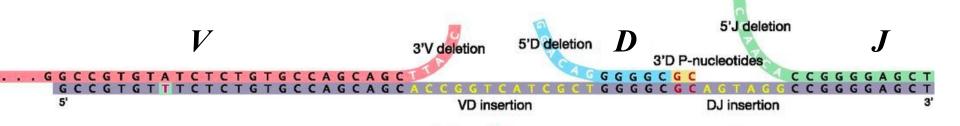
Why do we need a VDJ probabilistic model?



To compute **VDJ scenario**, we need to:

- perform VDJ classification to find germline segments (well-studied problem)
- specify deletions in gene segments
- specify non-genomic insertions
- specify addition of palindromes

Why do we need a VDJ probabilistic model?

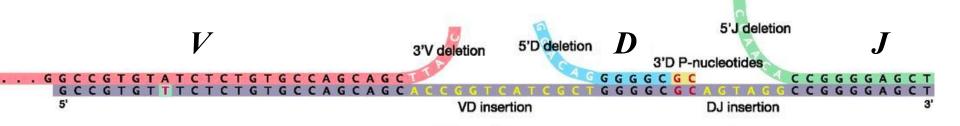


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Recombination events are not distributed uniformly

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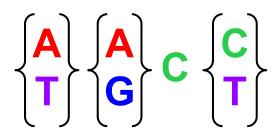
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Recombination events are not distributed uniformly

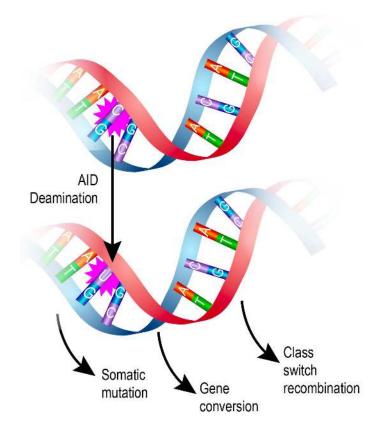
We need a probabilistic VDJ recombination model for a realistic description of these events

Why do we need an SHM probabilistic model?

SHM hotspots such as the degenerative 4-mers:



trigger mutations in antibodies



Somatic hypermutagenesis engages AID enzyme that changes immunoglobulin genes to improve antibody affinity

Building probabilistic SHM model

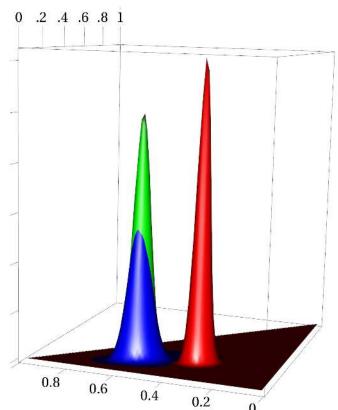
5-mer	Freq	A	С	G	T
ACAAC	83	-	0.24	0.48	0.28
GGCGT	1742	0.22	-	0.12	0.66
CCGTC	12	0.35	0.52	-	0.13
TCTCC	516	0.32	0.54	0.14	_

- The SHM model takes into account both the mutated nucleotide and its neighbours
- Detect new hot spots and compares SHMs in IG chains

Building probabilistic SHM model

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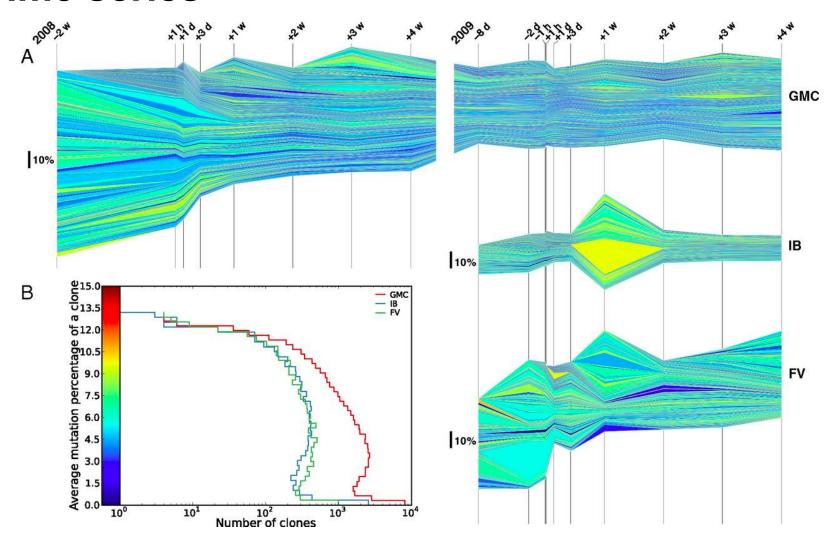


TCTCC 5-mer profiles for IGL, IGH, and IGK chains aggregated over 60 datasets

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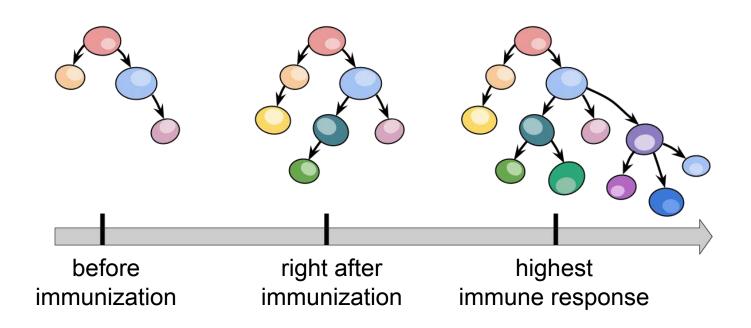
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Time series



Laserson et al, PNAS, 2014

Clonal analysis in time



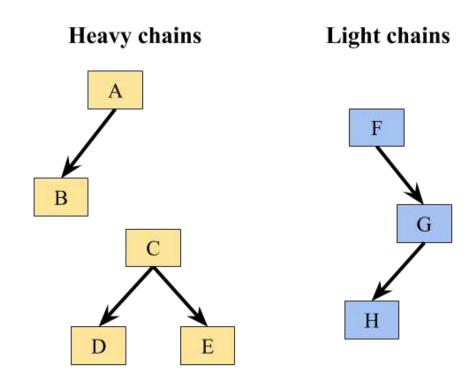
Clonal analysis of time series of antibody repertoire allows one to estimate efficiency of immune response



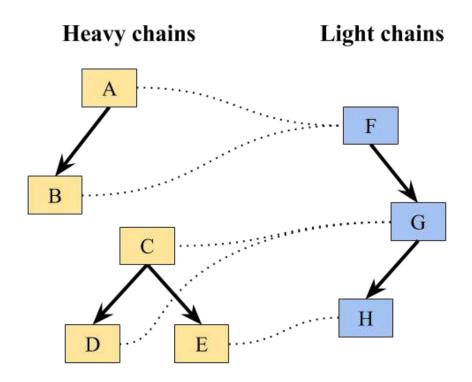
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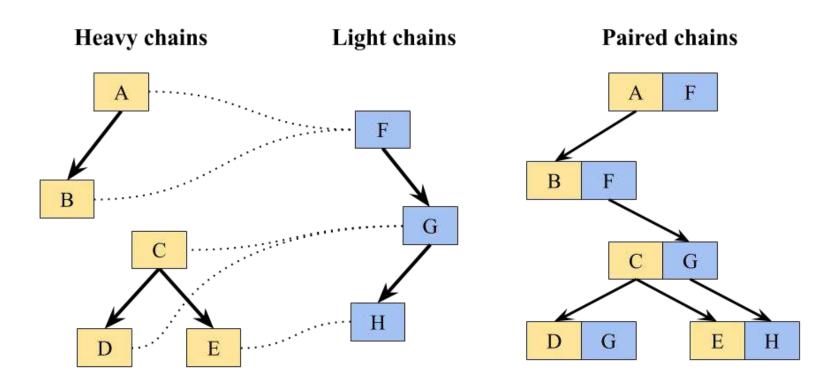
Clonal analysis for antibody repertoire



Clonal analysis for paired antibody repertoire



Clonal analysis for antibody repertoire

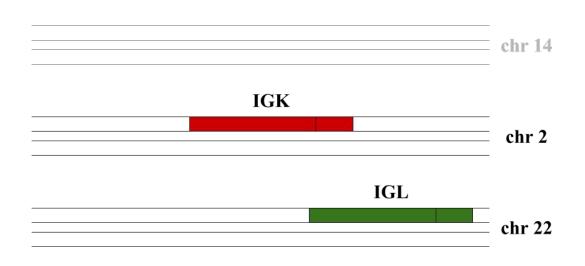


- utilizes information about chain pairing to construct paired clonal tree
- reveals that, contrary to previous views, B-cells often co-express multiple heavy and light chains.



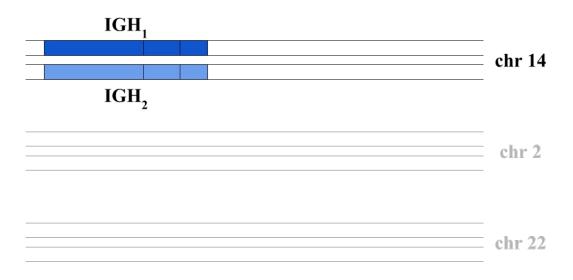
Light chain duality

co-expression of both kappa and lambda chains by a single Bcell



Allelic inclusion

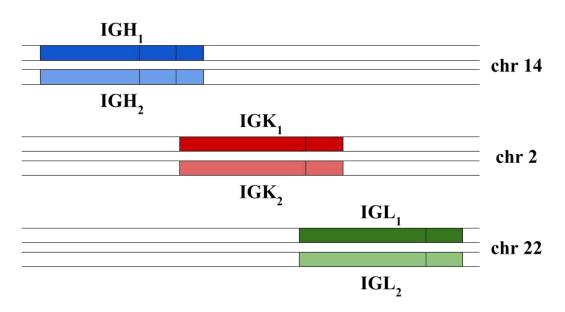
production of chains from both haplomes by B-cells



Casellas et al., *J Exp Med*, 2007 Beck-Engeser et al., *PNAS*, 1987

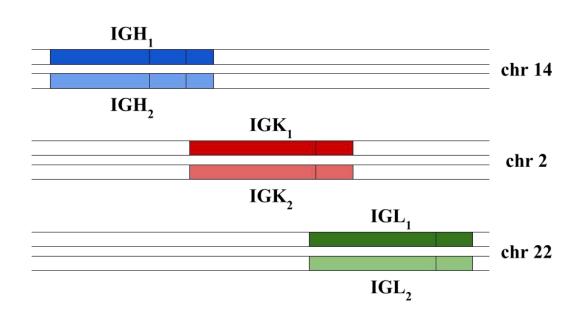
Duality + allelic inclusion

A single B-cell may express multiple chains due to allelic inclusions and/or light chain duality

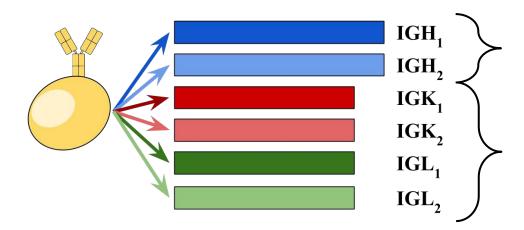


Multi-chain effect

A single B-cell may express multiple chains due to allelic inclusions and/or light chain duality

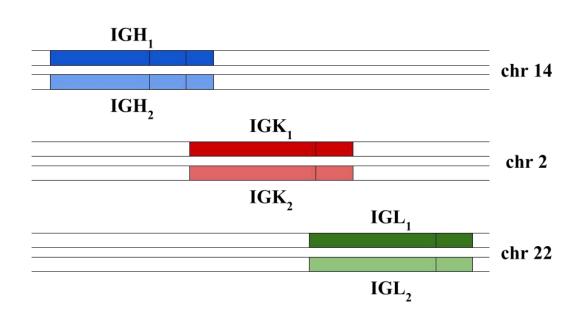


Multi-chain effect: B-cell can express up to 6 different chains:

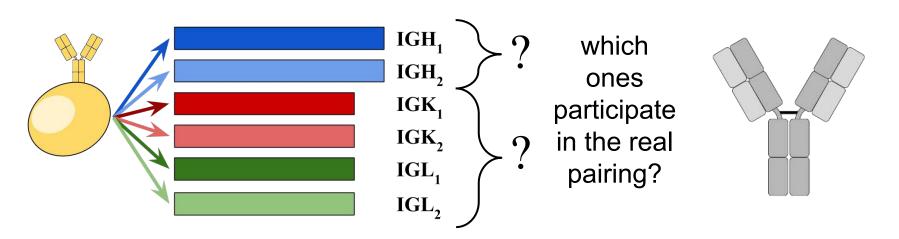


Multi-chain effect

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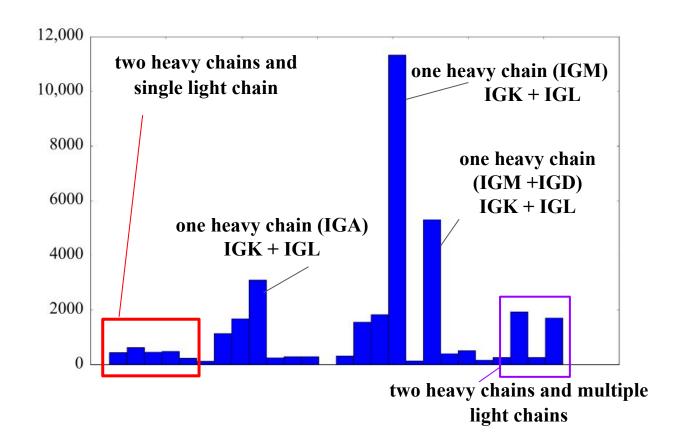


Multi-chain effect: B-cell can express up to 6 different chains:



Multi-chain effect is common in healthy B-cells!

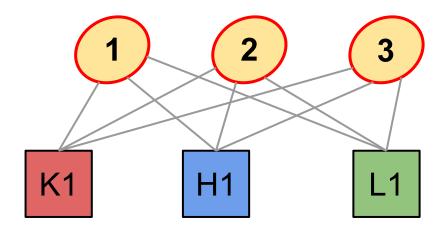
25% (!) of B-cells with known pairing have allelic inclusions and/or light chain duality



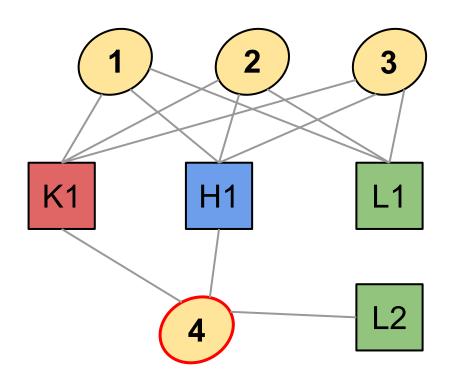
Cells 1, 2, and 3 express identical heavy, kappa and lambda chains. Thus, 1, 2, and 3 are clones of the same B-cell

Which light chain contributes to the antibody:

kappa or lambda?



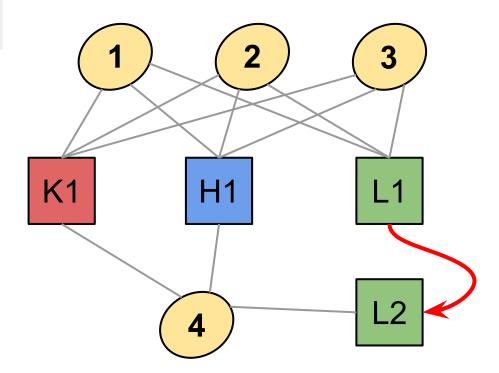
Cell 4 shares heavy and kappa chains with cells 1, 2 and 3, but has different lambda chain (L2)



Alignment of L1 and L2 reveals that L1 is an ancestor of L2

Thus, cell 4 is a descendant of cells 1, 2, and 3

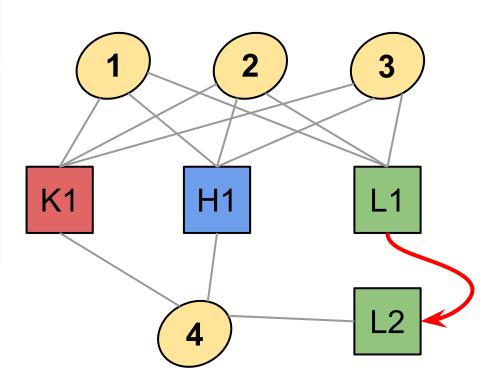
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Evolution of L1 into L2 provides evidence that cells 1, 2, 3, and 4 generate functional antibodies

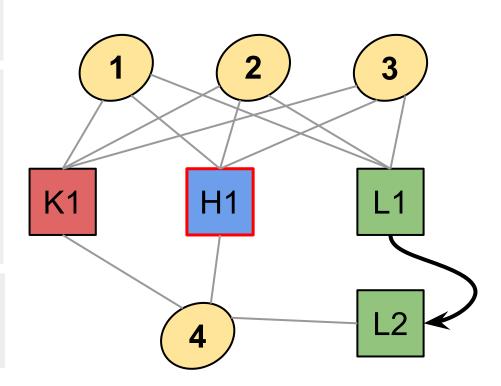


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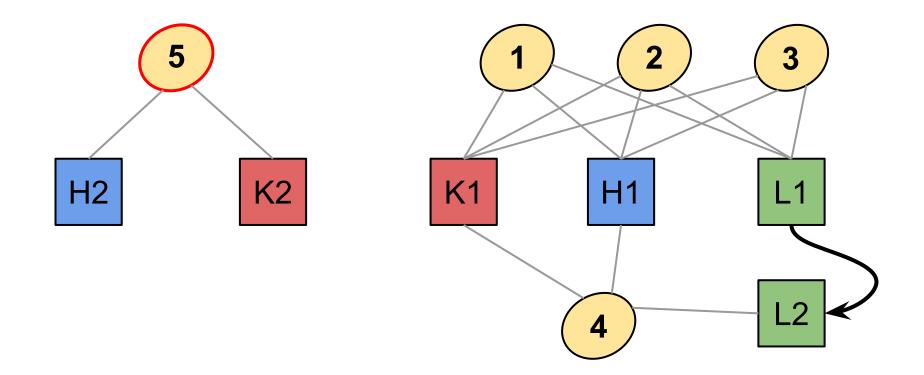
Evolution of L1 into L2 provides evidence that cells 1, 2, 3, and 4 generate functional antibodies

But it contradicts with a fact that H1 is non-productive



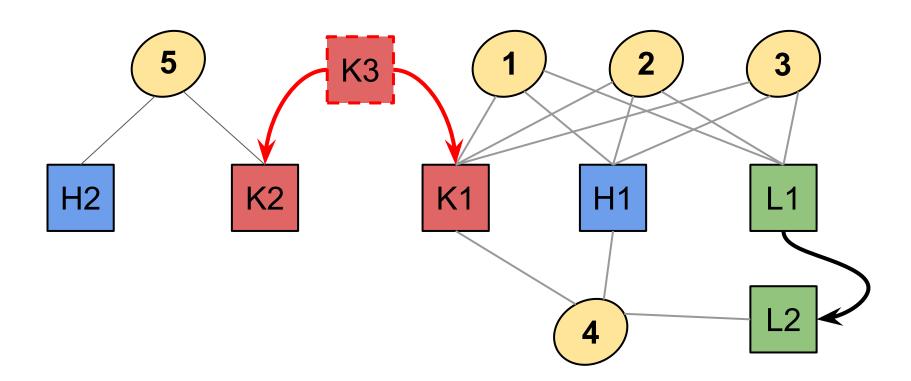
There are more B-cells to analyze!

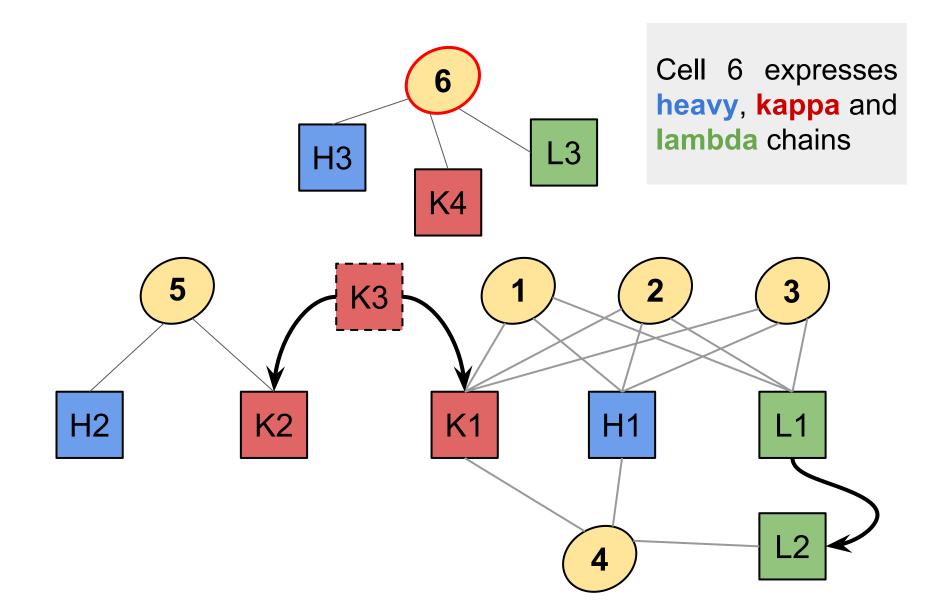
Cell 5 expresses heavy and kappa chains

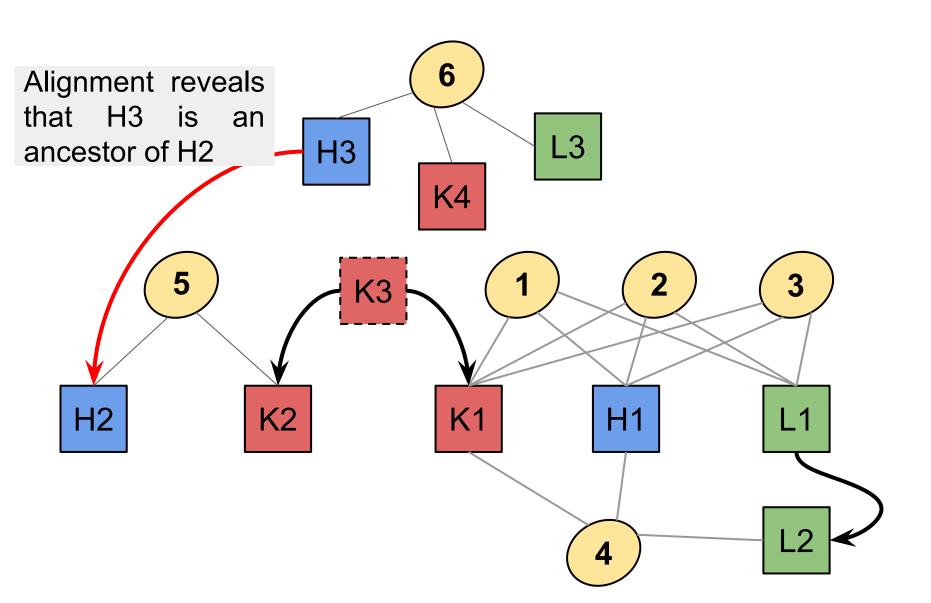


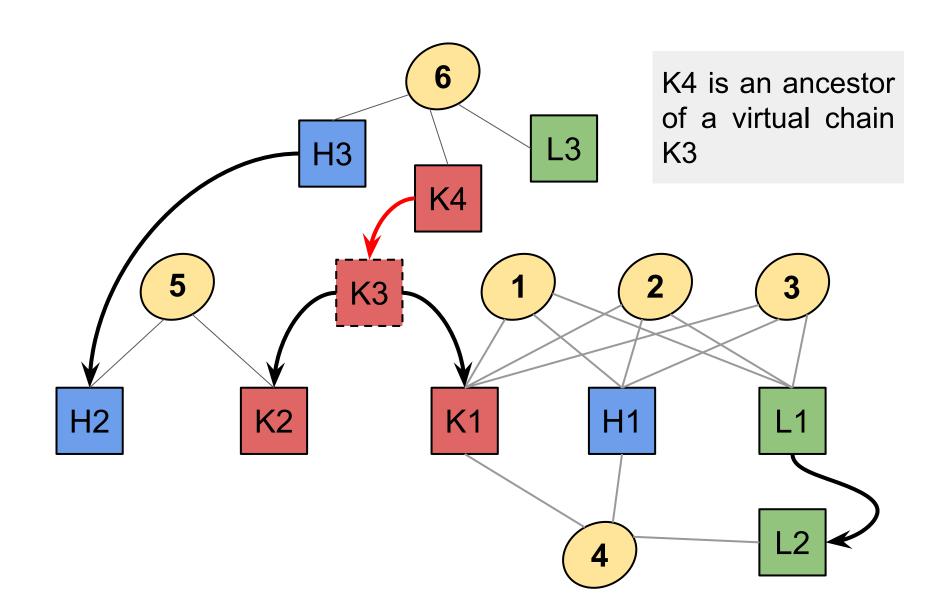
There are more B-cells to analyze!

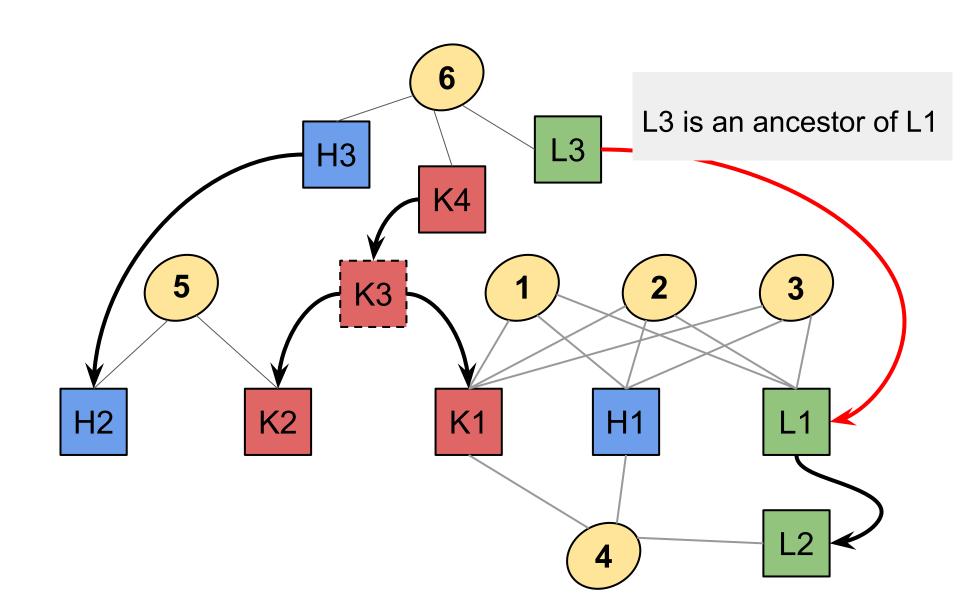
K2 and K1 have originated from a an unknown kappa chain K3 that is missing in the repertoire









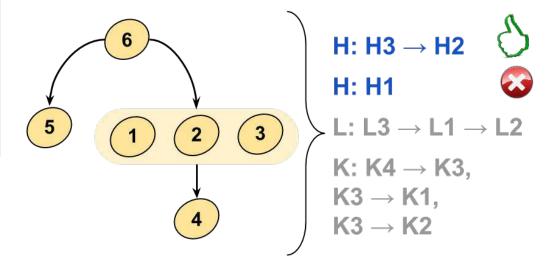


Evolutionary analysis helps to understand

true chain pairing

H1 lineage is nonproductive, so it does not participate in pairing 6 H3 K4 L3 K1 H1 L1 L1

Lineage H3 → H2 is more likely to participate in chain pairing



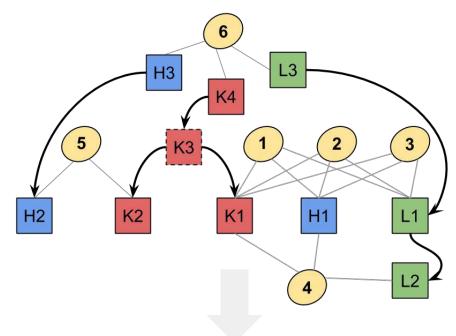
Evolutionary analysis helps to understand

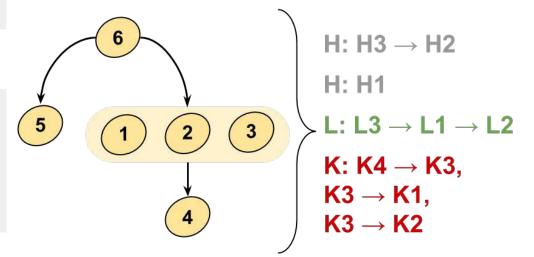
true chain pairing

 Lambda lineage contain synonymous mutations

- Mutations in lambda lineage are grouped into CDRs
- Mutations in kappa chain are distributed randomly along variable region

Lambda lineage undergoes selection, thus it more likely participates in chain pairing

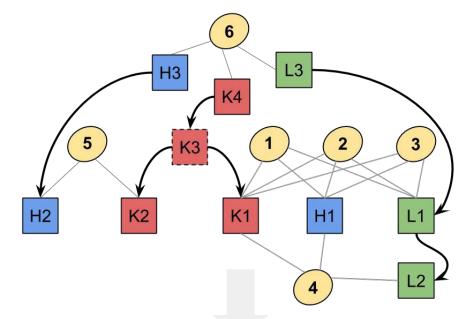


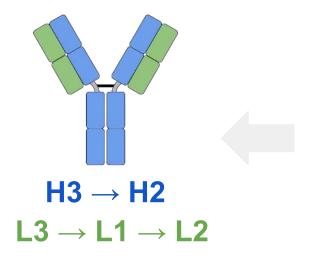


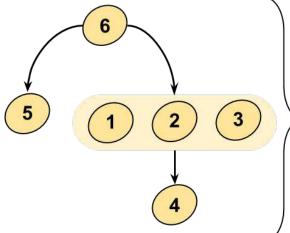
Evolutionary analysis helps to understand

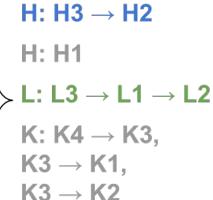
true chain pairing

Using information about clonal lineages for H, K and L chains and the SHM model, we can select the most likely chain pairing











Yana Safonova



Alexander Shlemov



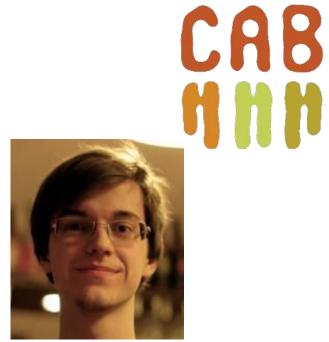
Andrey Bzikadze



Sergey Bankevich



Timofey Prodanov



Andrey Slabodkin



Alla Lapidus



Pavel A. Pevzner

Thank you!

